

HARMFUL ALGAE 2012



15차 국제 적조회의보고서

The 15th International Conference on Harmful Algae

October 29 - November 2, 2012, CECO,

Changwon, Gyeongnam, Korea

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INTERNATIONAL SOCIETY FOR THE STUDY OF HARMFUL ALGAE



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Aerial view of merry-go-round clay mitigation around fish cages in Yokjido Island, in the South Sea of Korea, August 21, 2008 (Photographed by Mr. JongSuk Jung)

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Growth response of *Pseudo-nitzschia circumpora* (Bacillariophyceae) to different salinities

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Abstract

Pennate diatoms from the genus *Pseudo-nitzschia* Peragallo are known to produce domoic acid and cause Amnesic Shellfish Poisoning (ASP). Although *Pseudo-nitzschia* spp. are commonly found in phytoplankton samples, no ASP has been documented in Malaysia to date. Clonal cultures of *Pseudo-nitzschia* were established and characterized using morphology through electron microscope for ultrastructural analysis. A newly described species, *Pseudo-nitzschia circumpora* was found in four locations in Malaysian waters, indicating the wide distribution of the species. In laboratory studies, *P. circumpora* from Malaysia showed a salinity tolerance from 25-35 psu, with an optimum growth at 30 psu. Further ecophysiological and toxinological studies are needed for a better knowledge of this newly described *Pseudo-nitzschia* species from Malaysia.

Keywords: *Pseudo-nitzschia circumpora*, morphology, physiology

Introduction

Research interests on marine diatom *Pseudo-nitzschia* have risen dramatically after it was confirmed as the causative organism for the first incidents of human intoxication in Prince Edward Island, 1987 (Subba Rao *et al.* 1988). The illness was later known as Amnesic Shellfish Poisoning (ASP) and the species responsible for the event was identified as *P. multiseriata* (Bates *et al.* 1989). ASP not only caused poisoning to human beings but also caused death of marine birds and marine mammals in subsequent years (Fritz *et al.* 1992; Scholin *et al.* 2000). Since then, the occurrence of *Pseudo-nitzschia* was well documented worldwide by various research groups (Lelong *et al.* 2012). Studies on the occurrence of *Pseudo-nitzschia* in Malaysia showed a high species diversity with 24 species recorded (Lim *et al.* 2012, 2013; Teng *et al.* 2013). One of these was found to produce high level of DA in cultures (Teng *et al.* 2014). In Malaysia, paralytic shellfish poisoning remained as the biggest concern for the seafood industry and public health due to blooms of the toxic dinoflagellates *Pyrodinium bahamense* (reviewed in Usup *et al.* 2012), *Alexandrium minutum* (Lim *et al.* 2004) and *Alexandrium tamiyavanichii* (Lim *et al.* 2004,

2006, 2007). Since 2009, studies were initiated to document the occurrence, distribution and genetic diversity of *Pseudo-nitzschia* species in order to assess the potential risk of ASP in Malaysian coastal waters. This contribution presents preliminary studies on the ecophysiology of *P. circumpora*.

Materials and Methods

Plankton samples were collected with a 20- μ m plankton net. Clonal cultures of *Pseudo-nitzschia* were established using SWII medium (Iwasaki 1961) at 30 psu and maintained under 25°C, 12:12 light: dark photoperiod with light intensity of approximately 100 μ mol photons m⁻² s⁻¹ in a cool-white fluorescence incubator (SHEL LAB, Cornelius, OR, USA). Natural and cultured materials were treated with acid for species identification under transmission electron microscope (TEM). *Pseudo-nitzschia circumpora* was cultured at different salinities ranging from 0-35 psu with sterilized SWII medium; cell densities were enumerated every two days to determine growth rates.

Results and Discussion

In the present study, the stability of morphological

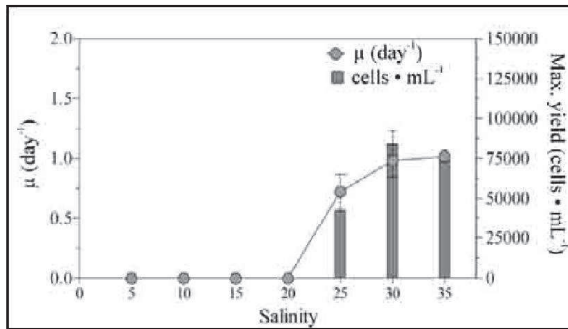


Figure 1. Growth rate, μ (d⁻¹) and maximum yield of *P. circumpora* strain PnSb58 under salinity treatments of 5 - 35 PSU.

characteristics of *P. circumpora* were examined from field samples and resulted with similar morphometric data. Morphometric comparison among the closely related species from the *P. pseudodelicatissima* complex showed that the number of poroids (in 1 μ m) and of dividing sectors are the most useful and distinctive morphological characteristics to discern *P. circumpora* from the others.

In terms of salinity tolerance, cell divisions were only observed within a salinity range of 25-35 psu (Fig. 1). This explains why *P. circumpora* can only be found in coastal waters of Malaysia but not in more brackish inner waters. No growth was recorded at salinity lower than 20 psu. Cell yield was highest (84,100 cell mL⁻¹) at 30 psu and lowest (<50,000 cells mL⁻¹) at 25 psu. The growth rate (μ) increased with increased salinities from 0.72 d⁻¹ at 25 psu to 1.01 d⁻¹ at 35 psu (Fig. 1).

The distribution of *P. circumpora* was documented. Only four out of seventeen sampling locations were recorded to have *P. circumpora* and these included: Port Dickson in Negeri Sembilan (Straits of Malacca), Sibulaut and Bintulu in Sarawak, and Semporna in Sabah.

Future ecophysiology and toxin production studies on this species are essential to enhance our understanding on this *Pseudo-nitzschia* species from Malaysian waters.

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